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| ***For Internal Use***  |
| **Accession/****Project No.** |  |
| **Date Rec’d** |  |
| **Initials** |  |



**Rutgers Translational Sciences**

**Research Pathology Services**

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| **PI** | Your PI *(Who is funding study?)* | **Contact Person** | Your name *(Who is submitting tissues?)* |
| **Institution** | Rutgers University | **Contact Email** | suzy.que@rutgers.edu |
| **Department** | Pharmacology and Toxicology | **Contact Phone** | 123-456-7890 |
| **PI Email** | joe.smith@rutgers.edu | **Date Submitted** | Day samples to RPS |

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| **Work Requested *(Check all that apply) (You may highlight instead of checking)*** |
| ☐ Wet Trimming  | ☐ Hematoxylin & Eosin Stain | ☐ Histopathology Report\* |
| ☐ Tissue Embedding | ☐ Histochemical Stains (List below)\*\* | ☐ Histopathology Scoring\* |
| ☐ Unstained Paraffin Sections | ☐ Immunohistochemistry (List below)\*\* | ☐ Slide scanning\*\*\* |

**\* Histopathology requires an additional form to be completed.**

**\*\* Histochemical staining and immunohistochemistry needs should be discussed with the Core Facility in advance.**

**\*\*\* Please indicate in “Notes” section p2 which slides to scan (if only selected slides being scanned)**

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| **Sample Information** |
| Total Number of Samples: 16\_\_\_  | Species: | Section Thickness:   5 um\_ |
|  **☐**  **Fixed Tissues**: \_\_**12**\_\_\_  | ☐ Mouse | Biosafety Level: |
|  ☐ Paraffin Blocks:            | ☐ Rat |  ☐ BL1 ☐ BL2 ☐ BL2+ |
|  **☐ Frozen Tissue:      4**  | ☐ Human  | IND-Enabling Study: |
|  ☐ Glass Slides:            |  ☐ Other:            |  ☐ Yes ☐ No |

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| **Project Information (Give a brief description of your project including any special instructions and information for histochemical stains, immunohistochemistry, etc)** |
| **Study Objective:** (What questions do you want histology to answer for you?)We want to know if “X” dosed rats had different microscopic findings than control rats in the following organs: liver, spinal cord, kidney, etc. The control rats are ID #s SD-1, L-2, F1 while the treated rats are ID #s L1, L3, SD-2 and F2. **Animal Identification:** What is age? what are genetic strains? dose groups? Previous findings (ie, studies with “X” dosed rats had clinical signs of front limb paralysis/paresis and histologic indications of unspecified cervical spinal cord abnormality. Multiple sections of brain and 25 to 40 different cervical, thoracic and lumbar spinal cord sections were examined.**Clinical / Gross observations:** -Did you note any behavioral abnormalities in any mice. If so, which one(s)? When you did the necropsies, did you notice anything in any mouse or mouse group that was different than controls? Any blood work/metabolic tests & results during the course of the study? (Glucose tolerance, Pyruvate tolerance, etc.)**Special instructions:** (Do your samples require orientation-specific embedding? Anatomically specific sectioning or sections at levels? Special stain request? If yes, please describe below.)Please add anything you like here. (ie, the *xxx* liver lobe is submitted for formalin processing/H&E; the *xxx* lobe was frozen in OTC compound/liquid nitrogen & reserved for possible Oil Red O special stain.Embed spinal cord for cross-sections. Take 3 sections at 100um levels. Longitudinal section of kidney, capturing adrenals where possible. Etc.) |

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| *For Internal Use* |
|  | Notification (Name and Date): |
| Initials: |

Upload form to iLab request and confirm sample drop off with:

Name: Marianne Polunas, RPh, PhD

Email: polunas@research.rutgers.edu

Address: 41 Gordon Rd. Suite B, Piscataway, NJ

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|  |  | **Individual Sample Information** |
| **RPS seq. #** | **PI’s Sample Identifier** | **Tissue(s)** **(i.e., skin, kidney)** | **# of H&E slides** | **# Un-stained slides** | **# Special Stained slides** | **# Sections (1 or 2?) *on Unstained*** | **Notes:** **ie, Slide scan (Y/N);****Other notes welcome** |
| **1** | **12-2021 55a** | **Liver** | **1** | **2** | **1**  | **2** | **Oil-red-O stain on frozen liver** |
| **2** | **12-2021 55b** | **Kidney** | **1** | **2** | **0** | **2** |  |
| **3** | **12-2021 55c** | **Spinal cord** | **1** | **2** | **0** | **3 levels at 100um** |  |
| **4** | **12-2021 56a** | **Liver** | **1** | **2** | **1**  | **2** | **Oil-red-O stain on frozen liver** |
| **5** | **12-2021 56b** | **Kidney** | **1** | **2** | **0** | **2** |  |
| **6** | **12-2021 56c** | **Spinal cord** | **1** | **2** | **0** | **3 levels at 100um** |  |
| **7** | **12-2021 57a** | **Liver** | **1** | **2** | **1**  | **2** | **Oil-red-O stain on frozen liver** |
| **8** | **12-2021 57b** | **Kidney** | **1** | **2** | **0** | **2** |  |
| **9** | **12-2021 57c** | **Spinal cord** | **1** | **2** | **0** | **3 levels at 100um** |  |
| **10** | **12-2021 58a** | **Liver** | **1** | **2** | **1**  | **2** | **Oil-red-O stain on frozen liver** |
| **11** | **12-2021 58b** | **Kidney** | **1** | **2** | **0** | **2** |  |
| **12** | **12-2021 58c** | **Spinal cord** | **1** | **2** | **0** | **3levels at 100um** |  |
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